PCT/EP 03/10494
Rec'd PCT/PTC 23 MAR 2005

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REC'D 28 OCT 2003
WIPO PCT

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Patentanmeldung Nr. | Patent application No. | Demande de brevet n°

02021597.6

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Europäisches Patentamt

European Patent Office

Office européen des brevets

Anmeldung Nr:
Application no.: 02021597.6
Demande no:

Anmeldetag:
Date of filing: 27.09.02
Date de dépôt:

Anmelder/Applicant(s)/Demandeur(s):

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SUISSE

Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:
(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.
If no title is shown please refer to the description.
Si aucun titre n'est indiqué se referer à la description.)

Microbial Production of Vitamin C from D-Sorbitol, L-Sorbose, L-Sorbosone or L-Gulose

In Anspruch genommene Prioriät(en) / Priority(ies) claimed /Priorité(s) revendiquée(s)
Staat/Tag/Aktenzeichen/State/Date/File no./Pays/Date/Numéro de dépôt:

Internationale Patentklassifikation/International Patent Classification/
Classification internationale des brevets:

C12P/

Am Anmeldetag benannte Vertragstaaten/Contracting states designated at date of filing/Etats contractants désignées lors du dépôt:

AT BE BG CH CY CZ DE DK EE ES FI FR GB GR IE IT LI LU MC NL PT SE SK TR

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Case 21418

Microbial Production of Vitamin C From D-Sorbitol, L-Sorbose, L-Sorbosone or L-Gulose

The present invention relates to the microbial production of L-ascorbic acid (vitamin C).

Vitamin C, which is one of very important and indispensable nutrient factors for human beings, has heretofore been commercially produced by the so-called "Reichstein method", which is well known as a technologically established process. This method, however, comprises a number of complex steps and any improvement in the overall yield is difficult to achieve. Therefore, there have been a number of proposals, which contemplate a reduction in the number of steps and/or an improvement in the overall yield.

Biosynthetic approaches for vitamin C production from D-glucose by cultivating plant tissues, algae, or yeast, which have key enzymes catalyzing L-galactono-γ-lactone or L-gulono-γ-lactone to vitamin C and the like are already known in the art. Moreover, the enzymatic reaction that catalyzes the oxidation of L-sorbosone to vitamin C is known. D-glucosone and L-sorbosone appear to be putative intermediates of vitamin C biosynthesis in detached bean and spinach leaves.

However, there have been no reports that bacteria produce vitamin C directly from any one of D-sorbitol, L-sorbose, L-sorbosone or L-gulose.

The present invention provides a process for the production of vitamin C from D-sorbitol, L-sorbose, L-sorbosone or L-gulose by bacteria, e.g. a microorganism selected from the strain *Gluconobacter oxydans* DSM 4025 (FERM BP-3812), a microorganism belonging to the genus *Gluconobacter* and having identifying characteristics of *G. oxydans* DSM 4025 (FERM BP-3812) and mutants thereof.

Hei/fm; 27.09.2002

More particularly the present invention provides a process for the production of vitamin C which process comprises culturing a microorganism selected from the strain *G. oxydans* DSM 4025 (FERM BP-3812), a microorganism belonging to the genus *Gluconobacter* and having identifying characteristics of *G. oxydans* DSM 4025 (FERM BP-3812) and mutants thereof, in an aqueous nutrient medium containing D-sorbitol, L-sorbose, L-sorbosone or L-gulose, and isolating and purifying vitamin C from the fermentation medium.

The present invention also provides a process for the production of vitamin C from D-sorbitol, L-sorbose, L-sorbosone or L-gulose which process comprises contacting a microorganism selected from the strain *G. oxydans* DSM 4025 (FERM BP-3812), a microorganism belonging to the genus *Gluconobacter* and having identifying characteristics of *G. oxydans* DSM 4025 (FERM BP-3812) and mutants thereof with D-sorbitol, L-sorbose, L-sorbosone or L-gulose in a reaction mixture and isolating and purifying vitamin C from the reaction mixture.

G. oxydans DSM 4025 was deposited at the Deutsche Sammlung von Mikroorganismen und Zellkulturen (DSMZ) in Göttingen (Germany), based on the stipulations of the Budapest Treaty, under DSM No. 4025 on March 17, 1987. The depositor was The Oriental Scientific Instruments Import and Export Corporation for Institute of Microbiology, Academia Sinica, 52 San-Li-He Rd., Beijing, Peoples Republic of China. The effective depositor was said Institute, of which the full address is The Institute of Microbiology, Academy of Sciences of China, Haidian, Zhongguancun, Beijing 100080, People's Republic of China.

Moreover, a subculture of the strain has also been deposited at the National Institute of Advanced Industrial Science and Technology (AIST), Tsukuba Central 6, 1-1-1 Higashi, Tsukuba, Ibaraki 305-8566, Japan, also based on the stipulations of the Budapest Treaty, under the deposit No. FERM BP-3812 on March 30, 1992. The depositor is Nippon Roche K.K., 6-1, Shiba 2-chome, Minato-ku, Tokyo 105-8532 Japan. This subculture may also be used in the present invention.

Mutants of *G. oxydans* DSM 4025 (FERM BP-3812) or a microorganism belonging to the genus *Gluconobacter* and having identifying characteristics of *G. oxydans* DSM 4025 (FERM BP-3812) may be obtained by treating the cells by means of, for instance, ultraviolet or X-ray irradiation, or a chemical mutagen such as nitrogen mustard or *N*-methyl-n′-nitro-*N*-nitrosoguanidine.

Any type of microorganism may be used, for instance, resting cells, acetone treated cells, lyophilized cells, immobilized cells and the like to act directly on the substrate. Any means

per se known as a method in connection with the incubation technique for micro-organisms may be adopted through the use of aeration and agitated submerged fermenters is particularly preferred. The preferred cell concentration range for carrying out the reaction is from about 0.01 g of wet cell weight per ml to 0.7 g of wet cell per ml, preferably from 0.03 g of wet cell per ml to 0.5 g of wet cell per ml.

The cultivation may be conducted at a pH of 4.0 to 9.0, wherein a pH value of about 5.0 to 8.0 may preferably be maintained. The cultivation period varies depending on the pH, temperature and nutrient medium to be used, and is preferably about 1 to 5 days, most preferably about 1 to 3 days. The preferred temperature range for carrying out the cultivation is from about 13°C to about 36°C, preferably from 18°C to 33°C. A preferred result may be obtainable from an incubation which utilizes a liquid broth medium.

As the nutrient medium for the incubation of the microorganism any aqueous nutrient medium including a carbon source, a nitrogen source, other inorganic salts, small amounts of other nutrients and the like, which can be utilized by the microorganism may be used. Various nutrient materials which are generally used for the better growth of microorganisms may suitably be included in the medium.

It is usually required that the culture medium contains such nutrients as assimilable carbon sources, for example glycerol, D-mannitol, erythritol, ribitol, xylitol, arabitol, inositol, dulcitol, D-ribose, D-fructose, D-glucose and sucrose in addition to the carbon sources converted to vitamin C; and digestible nitrogen sources such as organic substances, for example, peptone, yeast extract, baker's yeast, urea, amino acids and corn steep liquor. Various inorganic substances may also be used as nitrogen sources, for example nitrates and ammonium salts. Furthermore, the culture medium usually contains inorganic salts, for example magnesium sulfate, potassium phosphate and calcium carbonate.

For the advantageous performance of the incubation, any suitable factor which can promote the formation of the end product may be added to the medium.

Although the concentration of D-sorbitol, L-sorbose, L-sorbosone or L-gulose may also be varied with the cultivation conditions, a concentration of about 2 to 120 g/L is generally applicable, and wherein a concentration of 4 to 100 g/L is preferred.

The vitamin C thus produced and accumulated in the medium or reaction mixture may be separated and purified by any per se known conventional means which suitably utilized the property of the product, and it may be separated as the free acid or as a salt of sodium, potassium, calcium, ammonium or the like.

Specifically, the separation may be performed by any suitable combination or repetition of the following steps: by the formation of a salt, by using differences in properties between the product and the surrounding impurities, such as solubility, absorbability and distribution coefficient between the solvents, by absorption, for example on ion exchange resin. Any of these procedures alone or in combination constitutes a convenient means for isolating the product. The product thus obtained may further be purified in a conventional manner, e.g. by recrystallization or chromatography.

The identification of the vitamin C obtained by the method of this invention may be performed by, for instance, elemental anlysis as well as measurement of physicochemical properties such as spectrum of infrared absorption, mass spectrum, NMR and the like.

According to the present invention, the improvement in terms of the reduction in the number of steps is very significant because it leads to a one step pathway directed to the production of the vitamin C from any one of substrates D-sorbitol, L-sorbose, L-sorbosone or L-gulose.

In the following Examples, the process of the present invention will be illustrated in more detail.

Example 1: Conversion of D-sorbitol to vitamin C

One loopful of *G. oxydans* DSM 4025 (FERM BP-3812) grown on the agar medium containing 5.0 % D-mannitol, 0.25 % $MgSO_4 \cdot 7H_2O$, 1.75 % corn steep liquor, 5.0 % baker's yeast, 0.5 % urea, 0.5 % $CaCO_3$ and 2.0 % agar, which was cultivated at 27°C for 4 days, was inoculated into 5 ml of seed culture medium containing 8% D-sorbitol, 5 % baker's yeast, 0.05 % glycerol, 0.25 % $MgSO_4 \cdot 7H_2O$, 1.75 % corn steep liquor, 0.5 % urea, 1.5 % $CaCO_3$ and one drop of antifoam in test tube, and then cultivated at 30°C with 240 rpm for 20 hours on a reciprocal shaker.

Three ml of the seed culture were transferred into 500 ml Erlenmeyer flasks containing 50 ml of the production medium containing 8.0 % D-sorbitol, 5 % baker's yeast, 0.05 % glycerol, 0.25 % $MgSO_4 \cdot 7H_2O$, 3.0 % corn steep liquor, 1.5 % $CaCO_3$ and 0.15 % antifoam. The cultivation was carried out at 30°C with 180 rpm for 45 hours on a rotary shaker. Then, the concentration of vitamin C produced was measured by HPLC at a wavelength of 264 nm with the system which was composed of a UV detector (TOSOH UV8000; TOSOH Co., Kyobashi 3-2-4, Chuo-ku, Tokyo, Japan), a dualpump (TOSOH CCPE; TOSOH Co.), an integrator (Shimadzu C-R6A; Shimadzu Co., Kuwahara-cho 1, Nishinokyo, Chukyo-ku, Kyoto, Japan) and a column (YMC-Pack polyamine II; YMC,

Inc., 3233 Burnt Mill Drive Wilimington, NC 28403, USA), As a result, 118.1 mg/L of vitamin C was produced.

Example 2: Conversion of L-sorbose to vitamin C

One loopful of *G. oxydans* DSM 4025 (FERM BP-3812) grown on the agar medium containing 5.0 % D-mannitol, 0.25 % $MgSO_4 \cdot 7H_2O$, 1.75 % corn steep liquor, 5.0 % baker's yeast, 0.5 % urea, 0.5 % $CaCO_3$ and 2.0 % agar, which was cultivated at 27°C for 4 days, was inoculated into 5 ml of seed culture medium containing 8 % L-sorbose, 5 % baker's yeast, 0.05 % glycerol, 0.25 % $MgSO_4 \cdot 7H_2O$, 1.75 % corn steep liquor, 0.5 % urea, 1.5 % $CaCO_3$ and one drop of antifoam in test tube, and then cultivated at 30°C with 240 rpm for 20 hours on a reciprocal shaker.
Three ml of the seed culture were transferred into 500 ml Erlenmeyer flasks containing 50 ml of the production medium containing 8.0 % L-sorbose, 5 % baker's yeast, 0.05 % glycerol, 0.2 5% $MgSO_4 \cdot 7H_2O$, 3.0 % corn steep liquor, 1.5 % $CaCO_3$ and 0.15 % antifoam. The cultivation was carried out at 30°C with 180 rpm for 20 hours on a rotary shaker. As a result, 407.1 mg/L of vitamin C was produced.

Example 3: Production of vitamin C from D-sorbitol, L-sorbose, L-sorbosone and L-gulose with resting cell system

G. oxydans DSM 4025 (FERM BP-3812) was cultivated on the agar medium consisting of 8.0 % L-sorbose, 0.25 % $MgSO_4 \cdot 7H_2O$, 1.75 % corn steep liquor, 5.0 % baker's yeast; 0.5 % urea, 0.5 % $CaCO_3$ and 2.0 % agar at 27°C for 4 days. The cells of *G. oxydans* DSM 4025 (FERM BP-3812) grown on the above medium were transferred into 50 mM potassium phosphate buffer (pH 7.0) and washed twice with the same buffer. The optical density of the cell suspension at 600 nm was 21.9. It contained 0.057 g of wet cell weight per ml. The reaction mixture (5 ml in test tube) contained the cell suspension and 8 % D-sorbitol, 8 % L-sorbose, 0.5 % L-sorbosone or 1 % L-gulose in 50 mM potassium phosphate buffer (pH 7.0). The reaction was started by the inoculation of cell suspension and carried out at 30°C and with 180 rpm on a reciprocal shaker. The vitamin C content was measured at the reaction time of 4, 20 and 24 hours with HPLC. Table 1 shows the quantity of vitamin C produced from each substrate by *G. oxydans* DSM 4025 (FERM BP-3812).

Table 1:Vitamin C production from D-sorbitol, L-sorbose, L-sorbosone or L-gulose

Substrate	Vitamin C produced [mg/L]		
	4th hour	20th hour	24th hour
8% D-Sorbitol	0,0	62.3	90.3
8% L-Sorbose	636.1	908.0	874.3
0.5% L-Sorbosone	1,365.0	1,117.0	1,044.0
1% L-Gulose	488.8	1,355.0	1,673.0
None	0.0	0.0	0.0

Claims

1. A process for the production of vitamin C from D-sorbitol, L-sorbose, L-sorbosone or L-gulose by culturing a microorganism selected from the strain *Gluconobacter oxydans* DSM 4025 (FERM BP-3812), a microorganism belonging to the genus *Gluconobacter* and having identifying characteristics of *G. oxydans* DSM 4025 (FERM BP-3812) and mutants thereof, in an aqueous nutrient medium containing D-sorbitol, L-sorbose, L-sorbosone or L-gulose, and isolating and purifying vitamin C from the fermentation medium.

2. The process according to claim 1, wherein the culturing is carried out at pH values of about 4.0 to 9.0 and in a temperature range from about 13°C to 36°C for 1 to 5 days.

3. The process according to claim 1 or 2, wherein the culturing is carried out at pH values of about 5.0 to 8.0 and at a temperature range from about 18 to 33°C for 1 to 3 days.

4. A method for producing vitamin C from D-sorbitol, L-sorbose, L-sorbosone or L-gulose which comprises contacting a microorganism selected from the strain *Gluconobacter oxydans* DSM 4025 (FERM BP-3812), a microorganism belonging to the genus *Gluconobacter* and having identifying characteristics of *G. oxydans* DSM 4025 (FERM BP-3812) and mutants thereof with D-sorbitol, L-sorbose, L-sorbosone or L-gulose in a reaction mixture and isolating and purifying vitamin C from the reaction mixture.

5. The process according to claim 4, wherein the reaction is carried out at pH values of about 4.0 to 9.0 and at a temperature rang from about 13°C to 36°C for 1 to 48 hours.

6. The process according to claim 4 or 5, wherein the reaction is carried out at pH values of about 5.0 to 8.0 and at a temperature range from about 18°C to 33°C for 1 to 24 hours.

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PCT Application
EP0310494